METHOD FOR SCREENING FOR COMPOUNDS AS POTENTIAL SEDATIVES OR ANXIOLYTICS

5 The present invention relates to a method for screening a chemical compound for its potential as a sedative or anxiolytica. The invention also relates to a drug development method and to the use of a compound as identified by the screening method for the treatment, prevention or alleviation of anxiety, for inducing anaesthesia, pre-anaesthesia, muscle relaxation, or sedation, or for treatment, prevention or alleviation of fever cramps or status epilepticus in a subject.

10

BACKGROUND ART

GABA is the major inhibitory neurotransmitter in the mammalian brain and the $GABA_A$ receptor is the site of action of benzodiazepines. Multiple isoforms of $GABA_A$ 15 receptor exist; each receptor comprises a pentameric complex formed by co-assembly of subunits selected from 16 genes (α_{1-6}, β_{1-3}, γ_{1-3}, δ, ϵ, π, and θ) creating a chloride ion-channel.

The most abundant $GABA_A$ receptor in the mammalian brain comprises α, β, and γ subunits, and the classical anxiolytic benzodiazepines bind to these receptors if 20 they contain $\alpha_{1,2,3 \text{ or } 5}$ and γ_2 subunits. Because the subtypes are differently expressed in the brain as well as in other organs and because different subtypes are considered to be involved in different function, subtype specific compounds have been developed both with agonistic, antagonistic and inverse agonistic potentials. An example of such a subtype specific compound is the non-anxiolytic imidazopyridine zolpidem, which is 25 highly selective for α_1 containing $GABA_A$ receptors and is used as a short acting sedative in humans. α_2, α_3, and α_5 benzodiazepines sites are considered to be involved in anxiolytic properties and similar attempts have been made develop specific compounds for these sites. Such an example is the compound L-838,417, which is a selective α_2, α_3, and α_5 agonist [McKernan et al., Nat. Neurosci. 2000 June; 3(6); 587-30 92].

In order to develop new subtype specific compounds and to assess their efficacy in vivo, it is necessary to test new chemical entities (NCE's) in living animals. As the site of action is in the brain, behavioural testing is essential to determine pharmacokinetic and other ADME properties of the NCE. Furthermore, it is essential 35 to determine the efficacy in terms of hypnotic, sedative, anxiolytic, muscle relaxant, and anticonvulsive properties. Behavioural analyses in animals involve a number of so-called anxiety models, which detect the subjects' capability to take risks. The major problem with these models is that they are only partly predictive to assess a full behavioural response to a NCE with in vitro effect on the $GABA_A$ receptor. There

2

exists no *in vivo* prediction of alpha selectivity. Furthermore, because some of these compounds are sedative, it is hard to determine if their lack of action is specific or linked to its sedative properties. A method that activates systems in the brain relevant for the action of subtype specificity of NCE is therefore badly needed.

5   The hypothalamo-pituitary-adrenal (HPA) axis consists of the hypothalamic corticotrophin releasing factor (CRF) neurons in the medial parvocellular nuclei of the paraventricular nucleus (PVN), the corticotrophs of the anterior pituitary, and the steroid-producing cells in the adrenal cortex. The HPA axis drives the release of circulating corticosteroids in the blood, and is thus a central component of the stress

10 response. The HPA axis is under negative feedback, as increasing concentrations of plasma corticosteroids will inhibit the activity of the HPA axis via specific receptors for glucocorticosteroids. The HPA axis is under influence by other centers in the brain, and thereby it is activated in response to anxiety and fear. Pharmacological intervention can affect either directly on stress-related pathways, on the CRF neurons,

15 or peripherally to affect the inhibitory feedback on the axis.

Diazepam has been shown to slightly stimulate the HPA axis at the level of the hypothalamic corticotrophin releasing factor (CRF) neurons.

SUMMARY OF THE INVENTION

20

According to the invention it has now been found that activation of the HPA axis is coupled to mediation through the $GABA_A$ receptors comprising α_1-subtypes and thereby coupled to a sedative effect of the compound.

Thus, in a first aspect, the invention relates to a method for screening a chemical

25 compound for its potential as a sedative or anxiolytica, which method comprises the following steps:

a)   exposing the compound to a test system; and

b)   measuring the effect of the compound on the activity of the HPA axis.

In a second aspect, the invention relates to a drug development method

30 method, which comprises the identification of a compound by the screening method.

In a third aspect, the invention relates to the use of a compound identified in above method.

Other objects of the invention will be apparent to the person skilled in the art from the following detailed description and examples.

35

DETAILED DISCLOSURE OF THE INVENTION

In a first aspect, the invention provides a method for screening a chemical compound for its potential as a sedative or anxiolytica, which method comprises the

40 following steps:

3

a)   exposing the compound to a test system; and

b)   measuring the effect of the compound on the activity of the HPA axis.

In one embodiment, the chemical compound is a GABA$_A$ receptor modulator.

In a further embodiment, the test system is a test animal, and the compound is
5 exposed to the test animal by administration. In a still further embodiment, the test
animal is a non-human animal, such as a mammal. In a further embodiment, the test
animal is a rodent, such as a mouse or a rat. In a still further embodiment, the test
animal is a non-mammalian vertebrate, such as a reptile, bird or fish.

In a further embodiment, the route of administration of the compound is
10 intraperitoneal (i.p.), intraveneous (i.v.), peroral (p.o.) or subcutaneous (s.c.).

In a still further embodiment, the measurement of the activity of the HPA axis is
performed by measuring, in a blood sample from the test animal after administration,
the level of plasma corticosterone and/or ACTH.

In a still further embodiment, the test system is an explant system, such as
15 hypothalamic explant cultures, for example rat hypothalamic explant cultures.

In a further embodiment, the method for screening comprises the further step
of: c1) selecting the compound as a sedative drug candidate if the compound
substantially stimulates the HPA axis. In a special embodiment, the substantial
stimulation of the HPA axis is at least a 2-fold increase, preferably at least a 3-fold
20 increase, in corticosterone and/or ACTH over vehicle within the first two hours of
administration.

In a still further embodiment, the method for screening comprises the further
step of: c2) selecting the compound as an anxiolytica drug candidate if the compound
has substantially no effect on the HPA axis. In a special embodiment, the substantially
25 no effect on the HPA axis is less than a 50 percent increase, preferably less than a 25
percent increase, in corticosterone and/or ACTH over vehicle within the first two hours
of administration.

In a further aspect, invention provides a drug development method, which
comprises the identification of a compound according to the above method for
· 30 screening.

In a still further aspect, the invention provides the use of a compound identified
as a sedative drug candidate by the above method for screening or a
pharmaceutically acceptable salt thereof for the manufacture of a medicament for
inducing anaesthesia, pre-anaesthesia, muscle relaxation, or sedation, or for
35 treatment, prevention or alleviation of fever cramps or status epilepticus in a subject.

In a still further aspect, the invention provides the use of a compound identified
as an anxiolytic drug candidate by the above method for screening or a

4

pharmaceutically acceptable salt thereof for the manufacture of a medicament for the treatment, prevention or alleviation of anxiety.

In a further aspect, the invention provides a method for the treatment, prevention, or alleviation of anxiety comprising administering to said subject a
5 therapeutically effective amount of a compound identified as a antiolytica by the above method for screening or a pharmaceutically acceptable salt thereof.

In a still further aspect, the invention provides a method for inducing anaesthesia, pre-anaesthesia, muscle relaxation, or sedation, or for treatment, prevention or alleviation of fever cramps or status epilepticus anxiety comprising
10 administering to said subject a therapeutically effective amount of a compound identified as a sedative by the above method for screening or a pharmaceutically acceptable salt thereof.

Measurement of HPA axis activity

15   A good measure of the activity of the HPA axis (hypothalamus-pituitary-adrenal axis) is a measure of those hormones that are released in response to the activation, i.e. the adrenocorticotrophic hormone (ACTH) and glucocorticoids (such as corticosterone or cortisol). These hormones can easily be measured in the blood, urine and the saliva of the test animal. Furthermore, activation of the CRF neurons in
20 the hypothalamus can be assessed as activity of transcriptional activation in the neurons (Hoffman et al., J Neuroendocriol. 2002 Apr.; 14(4); 259-68).

One example of measuring the activity of the HPA axis is as follows: The animal is treated with the NCE and sacrificed within an hour. As the release of ACTH occurs within an hour after the stimulation of the CRF neurons, animals are sacrificed at t=0,
25 5, 15, 30 and 60 minutes after administration. Trunk blood is taken, serum separated and levels of ACTH is measured in the serum by specific radioimmunoassay. Similarly, the level of glucocorticosteroids are determined at t=0, 30, 60, and 120 minutes (the response occurs somewhat later than ACTH) using a radioimmunoassay.

30 **Pharmaceutical Compositions**

While a chemical compound as identified by the method according to the invention for use in therapy may be administered in the form of the raw chemical compound, it is preferred to introduce the active ingredient, optionally in the form of a physiologically acceptable salt, in a pharmaceutical composition together with one or
35 more adjuvants, excipients, carriers, buffers, diluents, and/or other customary pharmaceutical auxiliaries.

In a preferred embodiment, the invention provides pharmaceutical compositions comprising the chemical compound of the invention, or a pharmaceutically acceptable salt or derivative thereof, together with one or more pharmaceutically acceptable

5

carriers therefor, and, optionally, other therapeutic and/or prophylactic ingredients, know and used in the art. The carrier(s) must be "acceptable" in the sense of being compatible with the other ingredients of the formulation and not harmful to the recipient thereof.

5   The pharmaceutical composition of the invention may be administered by any convenient route which suit the desired therapy. Preferred routes of administration include oral administration, in particular in tablet, in capsule, in dragé, in powder, or in liquid form, and parenteral administration, in particular cutaneous, subcutaneous, intramuscular, or intravenous injection. The pharmaceutical composition may be
10 prepared by the skilled person using standard and conventional techniques appropriate to the desired formulation. When desired, compositions adapted to give sustained release of the active ingredient may be employed.

        Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing Co.,
15 Easton, PA).

BRIEF DESCRIPTION OF THE DRAWING

        The present invention is further illustrated by reference to the accompanying
20 drawing, in which:

        Fig. 1 shows the effect of increasing doses of zolpidem and L-838,417 on plasma corticosterone levels in mice. The data represent mean ± S.E.M. of 5 mice per group. Significant effect of compound compared to vehicle *p<0.05.

        Fig. 2 shows the time course of the effect of 10 mg/kg zolpidem on the HPA
25 axis. The rise in plasma ACTH precedes the rise in corticosterone.

        The following example will illustrate the invention further, however, it is not to be construed as limiting.

30   **EXAMPLES**

Example 1

Measuring the affect on the HPA axis of Zolpidem in mice
35   Adult male NMRI mice (23-27 g.) were purchased from Møllegaarden (Denmark). The animals were received at the animal facility, and housed 5 per cage under 12:12 light: dark cycle, humidity and temperature controlled room for at least 7 days before the experiment. Food and water were available ad libitum. All procedures were conducted in accordance with the Danish National Guide for Care and Use of
40 Laboratory animals. Zolpidem was purchased from Tocris Ltd (Bristol, UK) and L-

6

838,417 synthesised according to WO 98/04559 and was injected in a volume of 10 ml/kg and dissolved in 5% Chremophor.

The two drugs were administered (i.p.) at doses 0,025, 1,25, 2.5, 12.5 and 25 mg/kg. The mice were returned to their home cages and sacrificed by decapitation 60
5   minutes after drug administration and trunk blood was collected in centrifuge tubes containing 2 mg EDTA. Plasma aliquots were stored at –20°C until hormone levels were determined.

Plasma corticosterone was measured directly without prior extraction by a commercially [^{125}I] corticosterone radioimmunoassay kit from Amersham. The
10 experiment was performed twice. The data were analysed by a two-way analysis of variance (ANOVA) followed by the Dunn's test. All data are represented as group means and the standard error of means (SEM).

Zolpidem significantly and dose-dependently increased plasma corticosterone in doses from 0,5 mg/kg. As demonstrated in Fig. 1 the effect reached a maximum at
15 12.5 mg/kg and not further increased by 25 mg/kg. In contrast, L-838,417 had no effect on corticosterone in doses up to 12.5 mg/kg (Fig. 1). When tested 2 h after administration of 12.5 mg/kg L-838,417 no effects on plasma corticosterone was observed.